



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : G01N 33/543, 21/77	A1	(11) International Publication Number: WO 95/16914 (43) International Publication Date: 22 June 1995 (22.06.95)
(21) International Application Number: PCT/GB94/02741 (22) International Filing Date: 15 December 1994 (15.12.94) (30) Priority Data: 9325718.6 16 December 1993 (16.12.93) GB (71) Applicant (for all designated States except US): APPLIED RESEARCH SYSTEMS ARS HOLDING N.V. [NL/NL]; 6 John B. Gorsiraweg, Curacao (AN). (72) Inventors; and (75) Inventors/Applicants (for US only): ROBINSON, Grenville, Arthur [GB/GB]; 23 Burnham Way, Ealing, London W13 7YE (GB). FLETCHER, Janys [GB/GB]; 18 Devoil Close, Burpham, Guildford, Surrey GU21 5JY (GB). (74) Agent: FRANK B. DEHN & CO.; Imperial House, 15-19 Kingsway, London WC2B 6UZ (GB).		(81) Designated States: AU, CA, JP, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>With international search report.</i>
(54) Title: SENSOR DEVICE FOR SANDWICH ASSAY (57) Abstract Sensor devices for use in assaying for a ligand in a sample are described, the devices comprising: i) a discrete zone ("the measurement zone") on a region of which ("the measurement region") is immobilised directly or indirectly a first specific binding partner for the ligand under assay (or a reagent precomplexed with or capable of forming a complex with a specific binding partner for the ligand under assay), which zone additionally contains, in releasable form, a first known amount of an optionally labelled second specific binding partner for the ligand under assay, the second specific binding partner being directed to an epitope of the ligand under assay different to the epitope to which the first specific binding partner is directed; and ii) a second discrete zone ("the reference zone") on a region of which is immobilised directly or indirectly a first specific binding partner for the ligand under assay (or a reagent precomplexed with or capable of forming a complex with a specific binding partner for the ligand under assay), which zone additionally contains, in releasable form, a known amount of ligand analogue and separately contains, in releasable form, a second known amount of an optionally labelled second specific binding partner for the ligand under assay as defined above, said second known amount being less than the aforementioned first known amount in the measurement zone. Methods of sandwich assay using such devices are also described.		

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	GB	United Kingdom	MR	Mauritania
AU	Australia	GE	Georgia	MW	Malawi
BB	Barbados	GN	Guinea	NE	Niger
BE	Belgium	GR	Greece	NL	Netherlands
BF	Burkina Faso	HU	Hungary	NO	Norway
BG	Bulgaria	IE	Ireland	NZ	New Zealand
BJ	Benin	IT	Italy	PL	Poland
BR	Brazil	JP	Japan	PT	Portugal
BY	Belarus	KE	Kenya	RO	Romania
CA	Canada	KG	Kyrgyzstan	RU	Russian Federation
CF	Central African Republic	KP	Democratic People's Republic of Korea	SD	Sudan
CG	Congo	KR	Republic of Korea	SE	Sweden
CH	Switzerland	KZ	Kazakhstan	SI	Slovenia
CI	Côte d'Ivoire	LI	Liechtenstein	SK	Slovakia
CM	Cameroon	LK	Sri Lanka	SN	Senegal
CN	China	LU	Luxembourg	TD	Chad
CS	Czechoslovakia	LV	Latvia	TG	Togo
CZ	Czech Republic	MC	Monaco	TJ	Tajikistan
DE	Germany	MD	Republic of Moldova	TT	Trinidad and Tobago
DK	Denmark	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	US	United States of America
FI	Finland	MN	Mongolia	UZ	Uzbekistan
FR	France			VN	Viet Nam
GA	Gabon				

Sensor device for sandwich assay

5 The present invention relates to a method of conducting sandwich assays of chemical, biochemical or biological entities and to devices for use in such a method.

10 There is a now a great interest in the development of assay devices and techniques for the detection and measurement of the presence of an analyte in a sample, and the various methods and devices available have been extensively reviewed, for example in Biosensors: Fundamentals and Applications, edited by A.P.F. Turner, 15 I. Karube, G.S. Wilson, Oxford Scientific Publications, 1987. Standard assay techniques, however, are highly sensitive to a wide variety of conditions and interfering factors which may affect the level of the signal observed e.g. temperature, reagent stability, 20 incubation and development time. Accordingly, the analytical performance of standard assay techniques is often limited by the method of calibration of the immunosensor used, which usually involves carrying out an assay on a standard sample containing a known amount 25 of analyte. In respect of assays which involve an antibody, the immunological binding reactions which occur are frequently irreversible. Thus any calibration steps need to be carried out using a separate device or devices (preferably from the same manufacturing batch) 30 which inevitably introduces errors.

 The need for a separate calibration step involving the use of additional sensing devices can be avoided by using in the assay a device which is provided with separate zones whereby the calibration step is effected 35 within the assay procedure. Such methods applicable to both sandwich assays and competition assays are described in WO92/09892.

 Standard sandwich assay techniques are particularly

- 2 -

liable to exhibit the high dose 'hook' effect (a paradoxical reversal of the standard curve at high doses of analyte), a typical standard curve for a conventional immunoassay being illustrated in Figure 1. This effect
5 can be eliminated by using sequential rather than simultaneous application of the different specific binding partners for the analyte under assay. Alternatively, elimination of the high dose 'hook' effect necessitates assay of a sample at at least two
10 different dilutions. The most usual way, however, of performing a one-step sandwich assay is to employ a large excess of the labelled specific binding partner to alleviate the high dose 'hook' effect.

An alternative to using excess labelled specific
15 binding partner is to reference the sandwich assay by dosing a known amount of the analyte under assay into the assay device. In the absence of the analyte of interest in the sample a fixed signal will be obtained, but when analyte is present the dose-response curve of
20 the reference assay will be offset compared with that of the test assay, the relative magnitude of the offset decreasing with increasing analyte concentration. Eventually, however, at high analyte concentration the offset becomes zero as the immobilised specific binding
25 partner is saturated with analyte. This is illustrated for an immunoassay in Figure 2 and clearly, therefore, such referencing techniques will not be satisfactory at all concentrations of analyte present in the sample but especially at high analyte concentration and do not
30 alleviate the high-dose hook effect.

In the sandwich assay techniques described in WO92/09892 one method of referencing employs a device having a reference zone containing an immobilised specific antibody to the antigen under assay, the
35 reference zone also containing a pre-complexed mixture of a labelled second specific antibody to the antigen under assay and the antigen under assay. Although such a complex is likely to be more stable than the separate

- 3 -

components it is, unfortunately, unable to mimic the performance of the second specific binding partner in the measurement zone of the device. There can be also be a further disadvantage with this type of referencing in that should the second specific antibody have degraded during manufacture or storage of the device, no information can be gained about its reactivity by interrogating the reference zone. We have now found that these disadvantages can be overcome if the specific antibody and antigen under assay are initially present in the device separately rather than being pre-complexed.

We have now developed an assay device and a method of assay suitable for sandwich assays which overcome the problems of the known assay techniques and which offer means for calibrating the assay as part of the assay procedure.

Thus, according to a first aspect of the present invention there is provided a sensor device which comprises i) a discrete zone ("the measurement zone") on a region of which ("the measurement region") is immobilised directly or indirectly a first specific binding partner for the ligand under assay (or a reagent precomplexed with or capable of forming a complex with a specific binding partner for the ligand under assay), which zone additionally contains, in releasable form, a first known amount of an optionally labelled second specific binding partner for the ligand under assay, the second specific binding partner being directed to an epitope of the ligand under assay different to the epitope to which the first specific binding partner is directed; and ii) a second discrete zone ("the reference zone") on a region of which is immobilised directly or indirectly a first specific binding partner for the ligand under assay (or a reagent precomplexed with or capable of forming a complex with a specific binding partner for the ligand under assay), which zone additionally contains, in releasable form, a known

- 4 -

amount of ligand analogue and separately contains, in releasable form, a second known amount of an optionally labelled second specific binding partner for the ligand under assay as defined above, said second known amount
5 being less than the aforementioned first known amount in the measurement zone.

The term "ligand analogue" is used to denote a species which is capable of binding to the same epitopic site of the same specific binding partner as the ligand
10 under assay, and includes inter alia within its scope a known amount of ligand under assay.

The known amount of optionally labelled specific binding partner in the reference zone must be less than that in the measurement zone in order that the reference
15 zone gives rise to a signal which is constant and independent of the amount of analyte present in the sample up to the high dose hook concentration for the assay.

The device according to the invention may also
20 contain one or more additional reference zone(s), each reference zone present containing different known amounts of optionally labelled specific binding partner.

According to a further aspect of the present invention there is provided a method of assay for a
25 ligand in a sample which comprises the steps of

- i) incubating the sample with a device according to the invention as hereinbefore defined;
- ii) monitoring the signal appropriate to the assay technique employed arising from the measurement zone (as
30 hereinbefore defined) of said device ("the assay signal");
- iii) simultaneously or sequentially to the said monitoring in ii), monitoring the signal appropriate to the assay technique employed arising from the reference
35 zone(s) (as hereinbefore defined) of said device ("the reference signal(s)"); and
- iv) comparing the reference signal(s) with the assay signal, thereby determining using an appropriate

- 5 -

algorithm whether and/or the extent to which the ligand under assay is present in the sample.

The incubation in step i) involves contacting the sample with the measurement zone of said device and
5 simultaneously or sequentially contacting the sample with the reference zone(s) of said device.

A wide variety of devices may be used to perform the method of the present invention, including, for example, dipstick or "test-strip" biosensors, a device
10 using a "sample flow-through" configuration or devices employing sample containment. Examples of biosensors which may be used in the method of the present invention include sensors involving surface plasmon resonance, resonant mirror techniques, piezoelectric and total
15 internal reflectance techniques.

However, a preferred device according to the present invention is a capillary fill device, especially a fluorescence capillary fill device, for example the type of device described in EP-A-171148 or in WO-
20 90/14590. Such capillary fill devices may be used singly or in a suitable holder such as described in WO-90/1830.

As described in EP-A-171148, a capillary fill device (hereinafter CFD) typically consists of two
25 plates of transparent material, e.g. glass, separated by a narrow gap or cavity. One plate acts as an optical waveguide and carries an immobilised reagent appropriate to the test to be carried out in the device. As described in WO-90/14590, the other transparent plate
30 can carry on its surface remote from the cavity a layer of light-absorbing or opaque material. For use in a sandwich assay, the immobilised reagent may, for example, be a specific binding partner to the ligand desired to be detected and either one of the plates may
35 carry a soluble reagent comprising a further specific binding partner to the ligand under assay labelled with a fluorescent dye (the ancillary reagent). When a sample is presented to one end of the CFD it is drawn

- 6 -

into the gap by capillary action and dissolves the ancillary reagent. Because the capillary gap is narrow (typically about 100 microns) the binding reaction will generally go to completion in a short time, possibly
5 less than 5 minutes depending on the sample matrix, type of assay and reagent affinities. In a sandwich immunoassay for an antigen, a sample antigen will form a sandwich complex with a fluorescently labelled antibody and an antibody immobilised on the waveguide. Thus, for
10 a sandwich immunoassay, the amount of fluorescently labelled antibody which becomes indirectly bound to the waveguide by virtue of complex formation will, in general, be directly proportional to the concentration of antigen in the sample.

15 The term "antigen" as used herein will be understood to include both antigenic species, (for example, proteins, bacteria, bacterial fragments, cells, cell fragments and viruses) and haptens which may be rendered antigenic under suitable conditions.

20 Thus, according to a preferred embodiment of the device according to the present invention we provide a specifically-reactive sample-collecting and testing device for use in a sandwich assay for a ligand, possessing a cavity having two zones I and II mutually
25 separated and each zone carrying a layer comprising, in releasable form, a reagent suitable for the desired assay, said surface being a surface of a first solid plate fashioned of transparent material, wherein the wall of the or each cavity opposite to said first plate
30 comprises a second plate fashioned of transparent material and adapted to act as a light transmissive waveguide, the second plate having on its surface adjacent the cavity two zones IV and V corresponding in orientation to the aforementioned zones I and II
35 respectively, each of zones IV and V carrying a layer comprising an immobilised reagent suitable for the desired assay. The first plate advantageously carries on its external face an opaque coating.

The arrangement of the aforementioned zones is such that zone I is paired together with zone IV and zone II is paired together with zone V, such that one of said pairs provides the measurement zone as defined
5 hereinbefore and the other pair provides the reference zone as defined hereinbefore.

As mentioned earlier, CFDs and other devices
10 according to the invention may, if desired, contain more than one reference zone; and may if desired contain multiple measurement zones enabling simultaneous or sequential assays for ligands in the same sample to be conducted. For example, the device could contain a measurement zone and two or three reference zones as
15 herein defined for the same assay, to improve the accuracy of the calibration of the assay.

Alternatively, the device could contain a first measurement zone and a reference zone as herein defined for one assay together with a further measurement zone for a different assay; the reference zone would also
20 serve as a calibration for the further measurement zone although such calibration would differ from that for the measurement zone.

The reagents carried in the zones on the first transparent plate may be contained within a dissoluble
25 layer of a suitable material. After deposition of the soluble reagent, a capping layer e.g. polyvinyl alcohol (PVA) may be placed upon the reagent, which capping layer delays the dissolution of the reagent for a few seconds after the addition of the sample to the device.
30 This is to prevent the reagents being washed from one zone to another thereby precluding an accurate assay. The cavity or cavities of the device are preferably of a dimension small enough to enable sample liquid to be drawn into the cavity by capillary action, although any
35 other method of filling said cavities may be employed. The zones on the first transparent plate and thereby the corresponding zones on the second transparent plate may be arranged either in tandem or in any other geometrical

- 8 -

arrangement which maintains the integrity of the zones.

The capillary fill devices according to the invention may be manufactured by methods broadly similar to those described in EP-A-171148. According to a
5 further aspect of the present invention we also provide a method of manufacturing specifically-reactive sample-testing devices as described hereinbefore, comprising the steps of

(a) forming an array of patches of suitable reagents,
10 carried by zones I and II as described hereinbefore on the surface of a sheet material which is to provide part of a multiplicity of the devices,

(b) forming an array of patches of suitable reagents, carried by zones IV and V as described hereinbefore on
15 the surface of an additional structure, involving, where appropriate the immobilisation of specifically reactive species as described hereinbefore, said additional structure together with the said sheet material providing for each of the multiplicity of devices a
20 cavity for collecting and retaining a volume of sample liquid in contact with the said layers of suitable reagents, the cavity preferably being of capillary dimension, and

(c) separating the sheet material into portions each
25 providing one or a plurality of the sample-collecting and testing devices.

In this process, the zones of reagents contained on the first plate may be continuous if the reagents contained in the zones are of an identical nature.
30 Alternatively, the zones of reagents contained on the first plate, like the zones of reagents contained on the second plate, may be divided into a pattern of discrete portions, for example as a two-dimensional array of patches. When such patches are formed, they can be
35 made, for example, by firstly forming a continuous layer and then removing portions thereof to leave the desired pattern of identical reagent patches. Alternatively the desired pattern of patches may be applied directly by

- 9 -

any conventional printing method (for example by ink-jet printing or screen-printing), such a technique being most applicable to embodiments where, for each of the aforementioned plates, the reagents contained in the zones on said plate are not identical in nature or else are very expensive and their usage has to be kept to a minimum. Ink-jet printing is the preferred method of applying the reagents.

The immobilisation of a specifically reactive species onto the surface of the cavity may be carried out directly or indirectly. For example, when the specifically reactive species is an antibody, indirect immobilisation may be effected by means of an anti-species antibody which is itself bound to the said surface. Alternatively, immobilisation may be effected by conjugating an antibody with biotin and complexing with avidin pre-immobilised on the said surface; or vice versa. A further example of indirect immobilisation involves conjugating fluorescein isothiocyanate (FITC) to the specific binding partner for the species under assay and immobilising anti-FITC antibody onto said surface. Direct immobilisation may be effected by activating the said surface by treatment with a suitable reagent (e.g. a silanisation reagent such as aminopropyltrimethoxy-silane) to which the antibody can be covalently coupled using an appropriate cross-linking reagent (e.g. glutaraldehyde or glycolaldehyde). Alternative techniques well-known to the man skilled in the art may be used for immobilization of the said coating. Haptens and antigens may be immobilised directly onto the surface of the cavity by using appropriate immobilisation chemistry. Alternatively these haptens and antigens may be conjugated to a protein e.g. poly-L-lysine and then immobilised via the protein onto the cavity surface using known methods.

The mode of operation of one embodiment of the method according to the invention will now be described in terms of an immunoassay of an antigenic species.

- 10 -

The assay signal is determined by contacting the sample with an immobilised first specific antibody to the antigen under assay and a fluorescently labelled second specific antibody to the antigen under assay. The amount of labelled antibody which becomes bound in the measurement region as a result of an immunological reaction can be determined by standard methods and this amount can be related to the amount of antigen in the sample. The calibration of the assay is carried out by contacting the sample with an immobilised first antibody to the antigen under assay and with a non-precomplexed combination of a known amount of labelled specific second antibody to the antigen under assay and a known amount of antigen. When there is no analyte present in the sample the reagents in the reference zone give rise to a signal resulting from the known amount of labelled antibody in the reference zone since all the labelled antibody will become bound to the baseplate as a result of the immunological reaction involving the known amount of antigen. As the analyte concentration in the sample increases the known amount of labelled antibody will still bind to the immobilised reagent in the reference zone resulting in a signal which remains equivalent to that which arises when no analyte is present because of the fixed amount of labelled antibody in the reference zone i.e. the signal arising from the reference zone will be independent of the total concentration of antigen in the device. When the concentration of antigen in the sample is equivalent to that which results in an assay signal of a value of the high-dose hook region of the assay curve some labelled antibody in the reference zone will bind to antigen bound to the immobilised antibody and some will bind to antigen which remains free in solution. Accordingly, this will result in a decrease in the signal arising from the reference zone enabling the user to know that the concentration of analyte present in the sample is greater than the high-dose hook concentration. This embodiment and the

- 11 -

resulting signals obtained are illustrated in Figure 3.

The sandwich assay method according to the present invention provides the following specific advantages:

- 5 i) The reagents used in the reference zone are equivalent to those used in the measurement zone and they will therefore perform in the same manner i.e. they will respond similarly in both zones to variations in their environment,
- 10 ii) In the reference zone the (labelled) specific binding partner and the ligand analogue are separate i.e. they are not pre-complexed. Accordingly, it is possible to gain information about the quality of the specific binding partner (and its label) i.e. information as to whether it has degraded during
15 manufacture or storage of the device by suitable interrogation of the reference zone.
- 20 iii) There is a means to warn the operator that the concentration of analyte in the sample is very high and that the sample may, for example, therefore need to be diluted and re-assayed, namely the decrease in the signal from the reference zone at high analyte concentration.
- 25 iv) Should there be endogenous interfering factors in the sample (e.g. for an immunoassay where one of the specific binding partners is a polyclonal antibody the presence of antibodies such as, for example, human anti-mouse antibodies (HAMA)) then the reference zone will be influenced in the same way as the measurement zone. The reference zone can therefore be used to either correct
30 for such interference or to alert the operator to its existence.

A further advantage arises where a plurality of reference zones are employed, each reference zone containing different amounts of labelled specific
35 binding partner and ligand analogue. The signals arising from each of the reference zones will correspond to different dose values on the measurement zone assay dose/response curve and this will therefore allow for

- 12 -

full quantification of the assay. In particular, the reference zones can be constructed to give information about the middle region of the assay curve, this region often being the most useful region for an assay
5 determination. An illustration of a CFD containing four reference zones, and the signals which would arise from these zones as related to that from the assay zone is illustrated in Figure 4.

Although the embodiments described hereinbefore
10 illustrate the method of the invention to be particularly applicable to immunoassays, and in the preferred embodiments of the invention the ligand is a antigen and the specific binding partner comprises an antibody to the said antigen, the invention is not to be
15 taken as limited to assays of antibodies or antigens. Examples of ligands which may be assayed by the method of the invention are given in Table 1 below, together with an indication of a suitable specific binding partner in each instance.

20 In Figures 1 to 4 the symbols illustrated denote the following entities:-

- Y - Capture antibody
- L - Labelled second antibody
- 25 ■ - Sample antigen
- - Antigen dosed into the assay device

Table 1

Ligand	Specific Binding Partner
antigen	specific antibody
antibody	antigen
hormone	hormone receptor
hormone receptor	hormone
polynucleotide strand	complementary polynucleotide strand
protein A	immunoglobulin
immunoglobulin	protein A
lectins	specific carbohydrate
specific carbohydrate of lectins	lectins

The method of the invention has very broad applicability but in particular may be used to assay: hormones, including peptide hormones (e.g. thyroid stimulating hormone (TSH), luteinizing hormone (LH), human chorionic gonadotrophin (hCG), follicle stimulating hormone (FSH), insulin and prolactin) or non-peptide hormones, or thyroid hormones), proteins (e.g. carcinoembryonic antigen (CEA) and alphafetoprotein (AFP)), sugars, toxins, vitamins, proteins, viruses such as influenza, para-influenza, adeno virus, hepatitis, respiratory and AIDS viruses, or microorganisms.

It will be understood that the term "antibody" used herein includes within its scope:

- (a) any of the various classes or sub-classes of immunoglobulin, e.g. IgG, IgA, IgM, or IgE derived from any of the animals conventionally used, e.g. sheep, rabbits, goats or mice,
- (b) monoclonal antibodies,
- (c) intact molecules or "fragments" of antibodies, monoclonal or polyclonal, the fragments being those which contain the binding region of the antibody,

- 14 -

i.e. fragments devoid of the Fc portion (e.g. Fab, Fab', F(ab')₂) or the so-called "half-molecule" fragments obtained by reductive cleavage of the disulphide bonds connecting the heavy chain components in the intact antibody,

- (d) antibodies or antibody fragments produced or modified by recombinant DNA techniques.

The method of preparation of fragments of antibodies is well known in the art and will not be described herein.

As indicated above, various reagents may be optionally labelled. As examples of species for use as labels in the method according to the present invention are included fluorophores, enzymes, high refractive index particles and other species well known to the man skilled in the art. Preferably, fluorophores are used as labels. Examples of suitable fluorophores include fluorescein and its derivatives (e.g. fluorescein isothiocyanate (FITC)), rhodamine and its derivatives (e.g. XRITC, TRAP, TRITC), lucifer yellow, 2,4-dinitrofluorobenzene, phenylisothiocyanate, dansyl chloride, phycobiliproteins (e.g. allophycocyanin and phycoerythrin) and indocyanins.

It is also be advantageous to provide for additional compensation for various factors in the assay system which may influence the level of signal observed. Current assay techniques are highly sensitive to temperature, reagent stability, incubation and development time and other conditions and interfering factors which may affect the level of signal observed. This additional compensation can be achieved, for example, by using an assay method as hereinbefore described in which additional separate calibration step(s) are carried out. In such a method a device is used which is provided with appropriate reagents disposed in one or more zone(s) (calibration zone(s)) separate from the measurement zone and reference zone. The concept of using calibration zones for such

- 15 -

compensation is described in detail in WO92/09892.

The use of such calibration step(s) will serve two main purposes, namely

- 5 i) to confirm that the various reagents used in the assay procedure are performing according to their specification, and ii) to define a certain concentration level within the sample under test, and thereby to compensate for background fluorescent levels, temperature and pH changes and other factors which may
10 alter the level of the observed signals.

Alternatively, an additional calibration zone may be employed for compensating for "edge effects" as described in International Patent Application No. PCT/GB93/01217.

- 15 Thus according to a further aspect of the present invention we provide a device for use in an assay in which one or more additional calibration step(s) are carried out being a device as hereinbefore defined additionally comprising one or more further discrete
20 zone(s) ("the calibration zone(s)") on a region of which is immobilised directly or indirectly a reagent ("the calibration reagent") appropriate to the assay technique employed, which zone may also contain appropriate ancillary reagents suitable for the desired assay.

- 25 Thus, according to a further aspect of the present invention there is provided a method of assay for a ligand in a sample as hereinbefore defined, additionally comprising the steps of

- 30 v) simultaneously or sequentially to the incubation in step i), incubating the sample, if desired together with one or more ancillary reagents, with the calibration zone(s) of a device as hereinbefore defined;
- vi) monitoring the signal(s) ("the calibration
35 signal(s)") appropriate to the assay technique employed arising from the calibration zone(s); and
- vii) subsequently comparing the calibration signal(s) to both the assay signal and the reference signal as

- 16 -

hereinbefore defined and, using an appropriate algorithm, the measure of the extent to which the ligand under assay is present in the sample, as derived from the assay signal and the reference signal, is thereby calibrated.

5 - Manufacture of devices possessing a plurality of reference zones and/or one or more calibration regions as described above may be carried out by an analogous method to that described hereinbefore for devices
10 possessing only zones I and II, by additionally forming the patch of suitable reagents in the further zone(s) on the surface of the sheet material and immobilising the suitable reagents in the further zone(s) on the surface of the additional structure.

15 Where more than one calibration and/or reference zone is present, the reagents on each will generally be chosen such that the signals arising from each zone are not identical. Such non-identical signals can arise where the signal arising from each zone is the same
20 function of the amount of ligand present in the sample. One example is where the calibration reagents in each calibration zone are the same but the amounts of ancillary reagent(s) which form a complex with the calibration reagents in each zone differ. Another
25 example is where the calibration reagents in each calibration zone each give rise to a signal without the need for an ancillary reagent and are present in differing amounts. If it is found, despite such a choice of calibration reagents that identical signals
30 arise, then device failure (e.g. due to extremes of sample pH, too high a sample background signal, reagent degradation or interference in the assay by a factor in the sample) is indicated and the assay can be rejected; this is a further advantage of the present
35 invention.

 Thus, the reagents used in the calibration zone(s) are chosen so as to give rise to a zero or non-zero signal for the purposes of additional calibration of the

- 17 -

assay. The term "zero signal" denotes the background signal for the assay concerned. The term "non-zero signal" is to be construed accordingly. In a sandwich assay the zero signal will be the signal obtained when
5 no analyte is present.

Various methods may be used to calibrate the assay signal by means of the calibration signal(s). These methods can be summarised as either an additive, multiplicative or a combined additive/multiplicative
10 method. All methods rely on characterisation of the calibration region(s) during manufacture, so that any difference measured at the time of assay can be used to correct the data from the measurement region.

In a sandwich assay according to a further
15 embodiment of the present invention in which one or more additional calibration step(s) are carried out as hereinbefore described, in step v) either a) the calibration reagent (or optionally an ancillary reagent
precomplexed with or capable of forming a complex
20 involving the calibration reagent) is a specific binding partner for the ligand under assay, a labelled specific binding partner for the ligand under assay is present as an ancillary reagent and a known amount of the ligand under assay precomplexed to its labelled specific
25 binding partner is present as a yet further ancillary reagent or b) a labelled specific binding partner for the ligand under assay is present as an ancillary reagent and the calibration reagent (or optionally an ancillary reagent precomplexed with or capable of
30 forming a complex involving the calibration reagent) is a known amount of the ligand under assay precomplexed to its immobilized specific binding partner or c) a ligand distinct from the ligand under assay is present as an ancillary reagent and the calibration reagent (or
35 optionally an ancillary reagent precomplexed with or capable of forming a complex involving the calibration reagent) is a labelled specific binding partner for the ligand distinct from the ligand under assay or d) the

- 18 -

calibration reagent is a labelled binding partner non-specific for any ancillary reagent(s) present or e) the calibration reagent gives rise to the desired signal without the need for the presence of an ancillary
5 reagent.

A wide range of possibilities present themselves for the configuration of the calibration regions for use in the method of the present invention. These possibilities are set out in detail in W092/09892
10 incorporated herein by reference.

Thus according to a further aspect of the present invention we provide a device for use in an assay in which one or more additional calibration step(s) are carried out as hereinbefore described, being a
15 specifically-reactive sample-collecting and testing device as defined hereinbefore additionally carrying on said first plate one or more further zone(s) carrying a layer comprising, in soluble releasable form, ancillary reagent(s) suitable for the desired assay and
20 additionally carrying on said second plate one or more further zone(s) each of which is corresponding in orientation to one of said further zone(s) on said first plate, and each of which is carrying a layer comprising an immobilised calibration reagent as hereinbefore
25 defined.

The present invention further provides apparatus suitable for use in the method of assay according to the present invention as hereinbefore described which comprises a device according to the invention as
30 hereinbefore defined; and means for generating and monitoring the signals from the device.

- 19 -

Claims

1. A sensor device for use in a sandwich assay for a ligand in a sample which comprises i) a discrete zone ("the measurement zone") on a region of which ("the measurement region") is immobilised directly or indirectly a first specific binding partner for the ligand under assay (or a reagent precomplexed with or capable of forming a complex with a specific binding partner for the ligand under assay), which zone additionally contains, in releasable form, a first known amount of an optionally labelled second specific binding partner for the ligand under assay, the second specific binding partner being directed to an epitope of the ligand under assay different to the epitope to which the first specific binding partner is directed; and ii) a second discrete zone ("the reference zone") on a region of which is immobilised directly or indirectly a first specific binding partner for the ligand under assay (or a reagent precomplexed with or capable of forming a complex with a specific binding partner for the ligand under assay), which zone additionally contains, in releasable form, a known amount of ligand analogue and separately contains, in releasable form, a second known amount of an optionally labelled second specific binding partner for the ligand under assay as defined above, said second known amount being less than the aforementioned first known amount in the measurement zone.
2. A device as claimed in claim 1 also containing one or more additional reference zone(s), each reference zone present containing different known amounts of said optionally labelled second specific binding partner.
3. A device as claimed in claim 1 or claim 2 wherein the device is a capillary-fill device.
4. A device for use in a sandwich assay for a ligand

- 20 -

in a sample in which one or more additional calibration step(s) are carried out being a device as claimed in any one of claims 1 to 3 additionally comprising one or more further discrete zone(s) ("the calibration zone(s)") on
5 a region of which is immobilised directly or indirectly a reagent ("the calibration reagent") appropriate to the assay technique employed, which zone(s) may also contain appropriate ancillary reagents suitable for the desired assay.

10

5. A device as claimed in any one of claims 1 to 3 being a specifically-reactive sample-collecting and testing device for use in a sandwich assay for a ligand, possessing a cavity having two zones I and II mutually
15 separated and each zone carrying a layer comprising, in releasable form, a reagent suitable for the desired assay, said surface being a surface of a first solid plate fashioned of transparent material, wherein the wall of the or each cavity opposite to said first plate
20 comprises a second plate fashioned of transparent material and adapted to act as a light transmissive waveguide, the second plate having on its surface adjacent the cavity two zones IV and V corresponding in orientation to the aforementioned zones I and II
25 respectively, each of zones IV and V carrying a layer comprising an immobilised reagent suitable for the desired assay.

6. A device as claimed in claim 4, being a
30 specifically-reactive sample-collecting and testing device as claimed in claim 5 additionally carrying on said first plate one or more further zone(s) carrying a layer comprising, in soluble releasable form, ancillary reagent(s) suitable for the desired assay and
35 additionally carrying on said second plate one or more further zone(s) each of which is corresponding in orientation to one of said further zone(s) on said first plate, and each of which is carrying a layer comprising

- 21 -

an immobilised calibration reagent as defined in claim 4.

7. A device as claimed in claim 5 or claim 6 wherein the first plate carries on its surface remote from the cavity a layer of light-absorbing or opaque material.

8. A method of sandwich assay for a ligand in a sample which comprises the steps of

i) incubating the sample with a device as claimed in any one of claims 1 to 3, 5 or 7.

ii) monitoring the signal appropriate to the assay technique employed arising from the measurement zone (as hereinbefore defined) of said device ("the assay signal");

iii) simultaneously or sequentially to the said monitoring in ii), monitoring the signal appropriate to the assay technique employed arising from the reference zone(s) (as defined in claim 1) of said device ("the reference signal(s)"); and

iv) comparing the reference signal(s) with the assay signal, thereby determining using an appropriate algorithm whether and/or the extent to which the ligand under assay is present in the sample.

9. A method of sandwich assay for a ligand in a sample in which one or more additional calibration step(s) are carried out, being a method as claimed in claim 8 wherein in step i) the sample is incubated in the presence of a device as claimed in any one of claims 4, 6 or 7, additionally comprising the steps of

v) simultaneously or sequentially to the incubation in step i), incubating the sample, if desired together with one or more ancillary reagents, with the calibration zone(s) of a device as claimed in any one of claims 4, 6 or 7;

vi) monitoring the signal(s) ("the calibration signal(s)") appropriate to the assay technique

- 22 -

- employed arising from the calibration zone(s); and
vii) subsequently comparing the calibration signal(s) to both the assay signal and the reference signal as defined in claim 8 and, using an appropriate
5 algorithm, the measure of the extent to which the ligand under assay is present in the sample, as derived from the assay signal and the reference signal, is thereby calibrated.
- 10 10. A method as claimed in claim 9 wherein, in step v) either a) the calibration reagent (or optionally an ancillary reagent precomplexed with or capable of forming a complex involving the calibration reagent) is a specific binding partner for the ligand under assay, a
15 labelled specific binding partner for the ligand under assay is present as an ancillary reagent and a known amount of the ligand under assay precomplexed to its labelled specific binding partner is present as a yet further ancillary reagent or b) a labelled specific
20 binding partner for the ligand under assay is present as an ancillary reagent and the calibration reagent (or optionally an ancillary reagent precomplexed with or capable of forming a complex involving the calibration reagent) is a known amount of the ligand under assay
25 precomplexed to its immobilized specific binding partner or c) a ligand distinct from the ligand under assay is present as an ancillary reagent and the calibration reagent (or optionally an ancillary reagent precomplexed with or capable of forming a complex involving the
30 calibration reagent) is a labelled specific binding partner for the ligand distinct from the ligand under assay or d) the calibration reagent is a labelled binding partner non-specific for any ancillary reagent(s) present or e) the calibration reagent gives
35 rise to the desired signal without the need for the presence of an ancillary reagent.

11. A method of manufacturing a specifically-reactive

- 23 -

sample-testing device as claimed in claim 5, comprising the steps of

- 5 (a) forming an array of patches of suitable reagents, carried by zones I and II as defined in claim 5 on the surface of a sheet material which is to provide part of a multiplicity of the devices,
- 10 (b) forming an array of patches of suitable reagents, carried by zones IV and V as defined in claim 5 on the surface of an additional structure, involving, where appropriate the immobilisation of said reagents, said additional structure together with the said sheet material providing for each of the multiplicity of devices a cavity for collecting and retaining a volume of sample liquid in contact with the said layers of
- 15 suitable reagents, the cavity preferably being of capillary dimension, and
- (c) separating the sheet material into portions each providing one or a plurality of the sample-collecting and testing devices.

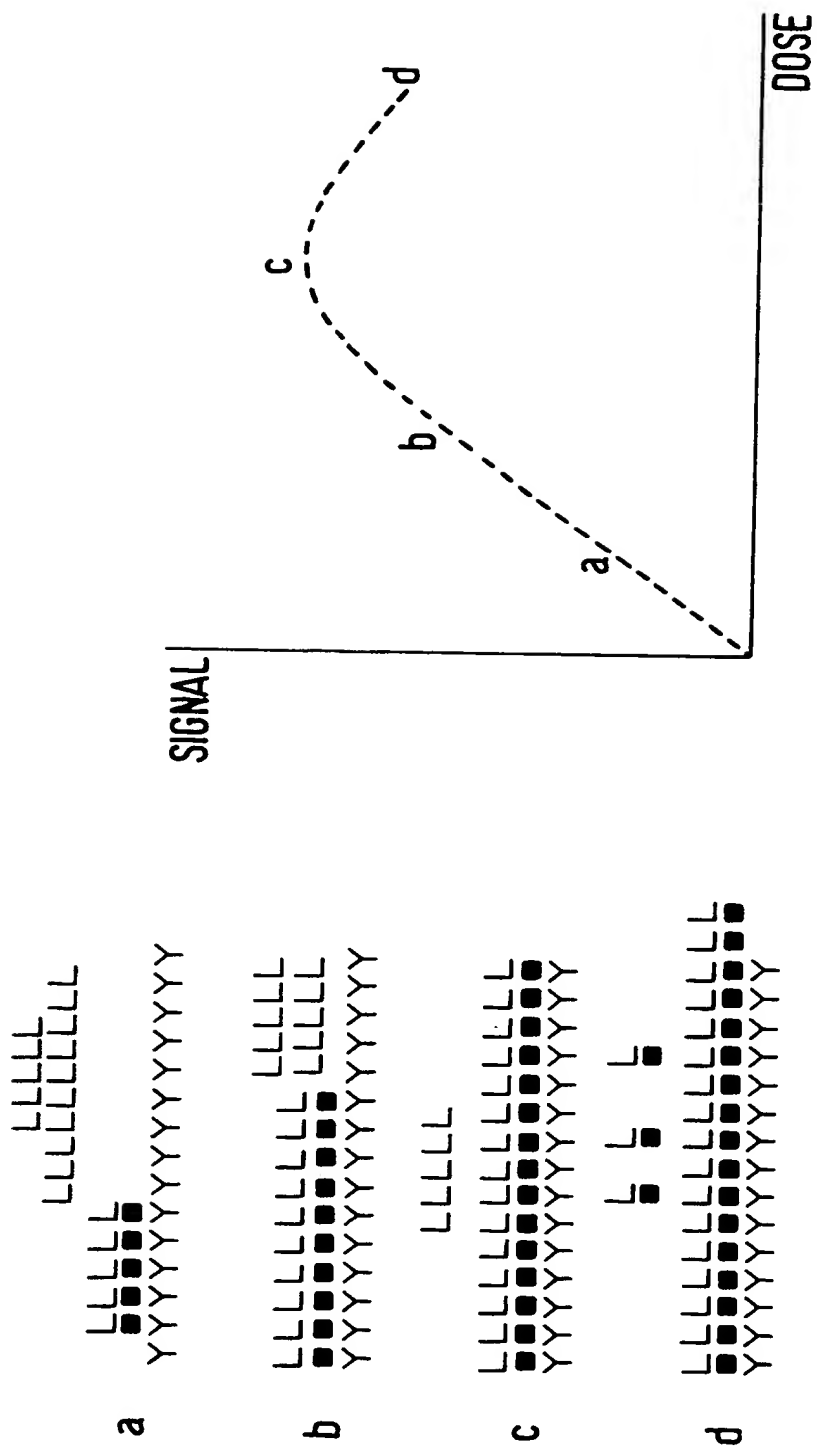
20

12. A method for the manufacture of a device as claimed in claim 6 which comprises a method as claimed in claim 11 additionally comprising the steps of forming the patch of suitable reagents in the further zone(s) on the
- 25 surface of the sheet material and immobilising the suitable reagents in the further zone(s) on the surface of the additional structure.

13. Apparatus suitable for use in a method of assay as
- 30 claimed in any one of claims 8 to 10 which comprises a device as claimed in any one of claims 1 to 7; and means for generating and monitoring the signals from the device.

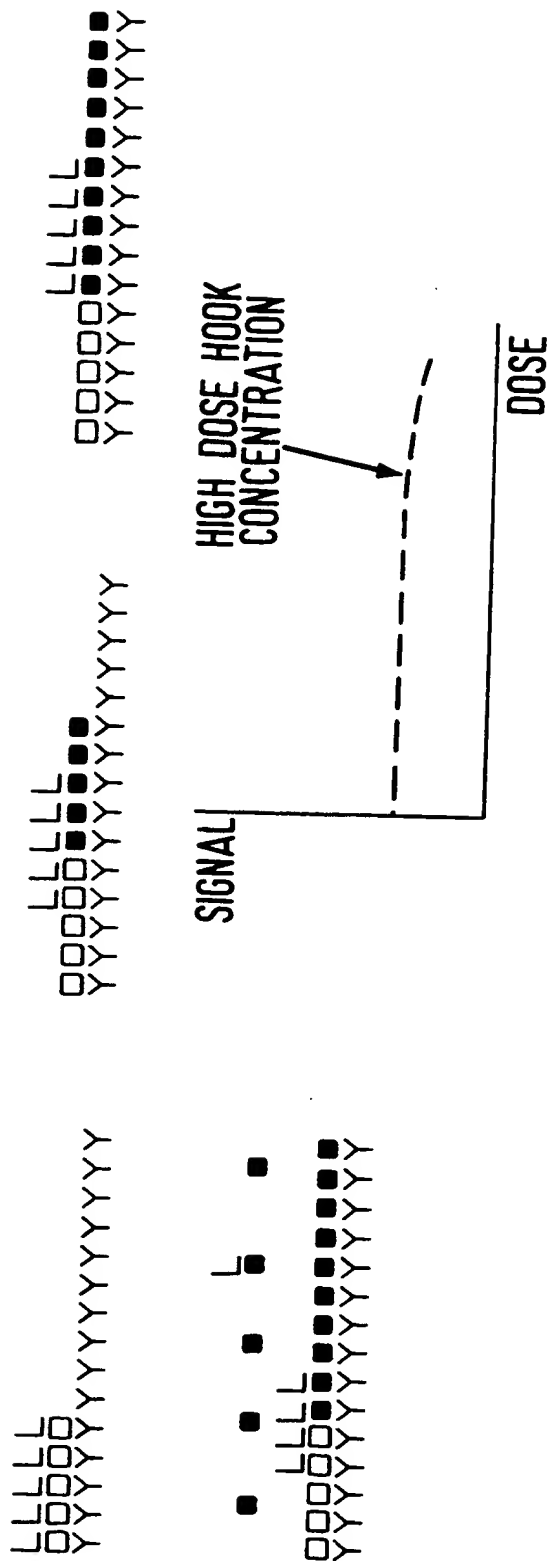
1/4

FIG. 1



3/4

FIG. 3



4 / 4

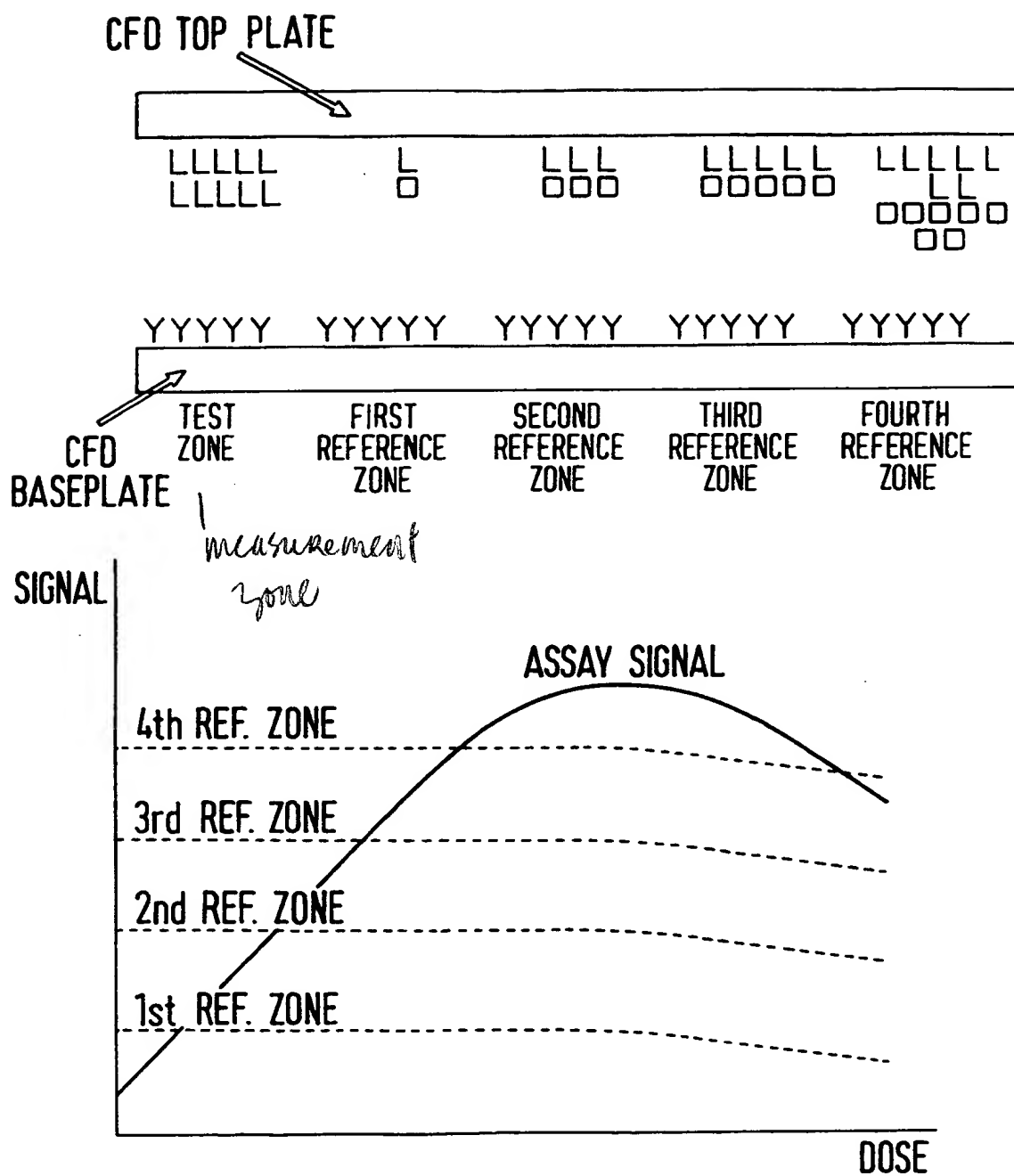


FIG. 4

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/GB 94/02741

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 G01N33/543 G01N21/77

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO,A,92 09892 (APPLIED RESEARCH SYSTEMS ARS HOLDING N.V.) 11 June 1992 cited in the application see claims ---	1-13
P,X	WO,A,93 25908 (ARS HOLDING 89 N.V.) 23 December 1993 see claims; examples ---	1-13
A	EP,A,0 422 708 (ARES-SERONO RESEARCH & DEVELOPMENT LIMITED PARTNERSHIP) 17 April 1991 cited in the application see the whole document --- -/--	1

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- "&" document member of the same patent family

Date of the actual completion of the international search

16 March 1995

Date of mailing of the international search report

29.03.95

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+ 31-70) 340-3016

Authorized officer

Moreno, C

INTERNATIONAL SEARCH REPORT

Int. onal Application No
PCT/GB 94/02741

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>WO,A,90 14590 (ARES-SERONO RESEARCH & DEVELOPMENT LIMITED PARTNERSHIP) 29 November 1990 cited in the application see the whole document -----</p>	1

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No
PCT/GB 94/02741

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A-9209892	11-06-92	AU-A- 8921691	25-06-92
		CA-A- 2095245	23-05-92
		EP-A- 0558680	08-09-93
		JP-T- 6502720	24-03-94
<hr/>			
WO-A-9325908	23-12-93	AU-B- 4343593	04-01-94
		CA-A- 2137654	23-12-93
<hr/>			
EP-A-0422708	17-04-91	AU-A- 2967289	25-05-89
		AU-B- 583040	20-04-89
		AU-A- 4491085	10-01-86
		AU-B- 588245	14-09-89
		AU-A- 4491185	10-01-86
		AU-B- 581669	02-03-89
		AU-A- 4491385	10-01-86
		CA-A- 1231136	05-01-88
		CA-A- 1246891	20-12-88
		CA-A- 1261256	26-09-89
		EP-A,B 0171148	12-02-86
		EP-A,B 0170375	05-02-86
		EP-A,B 0170376	05-02-86
		WO-A- 8600135	03-01-86
		WO-A- 8600141	03-01-86
		WO-A- 8600138	03-01-86
		JP-B- 3010902	14-02-91
		JP-T- 61502418	23-10-86
		JP-T- 61502419	23-10-86
		JP-B- 2024459	29-05-90
		JP-T- 61502420	23-10-86
		US-A- 4978503	18-12-90
		US-A- 4810658	07-03-89
<hr/>			
WO-A-9014590	29-11-90	AU-B- 617254	21-11-91
		AU-A- 5740390	18-12-90
		CA-A- 2031515	19-11-90
		DE-D- 69004049	25-11-93
		DE-T- 69004049	07-04-94
		EP-A- 0426817	15-05-91
		ES-T- 2045921	16-01-94
		JP-T- 3506078	26-12-91